

## SPECIALIA

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A new chalcone glucoside and cernuoside from the flowers of *Acacia dealbata*F. Imperato<sup>1</sup>

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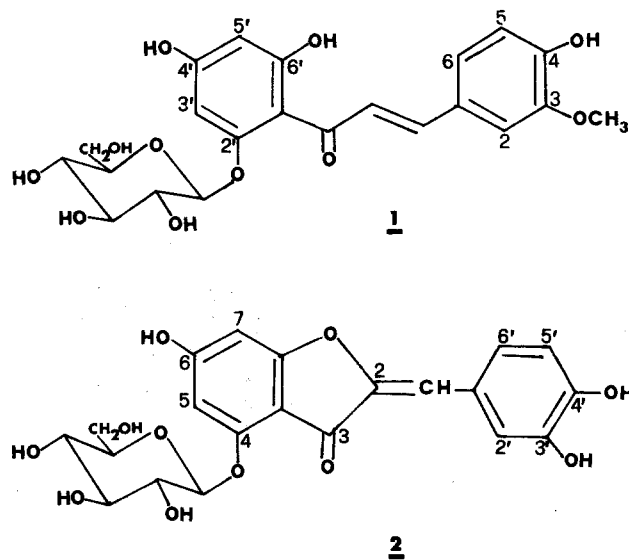
**Summary.** A new chalcone glucoside has been isolated from the flowers of *Acacia dealbata* and shown to be 4,2',4',6'-tetrahydroxy-3-methoxychalcone 2'-O- $\beta$ -D-glucoside (**1**) by chemical degradations and spectroscopic methods. Cernuoside (4,6,3',4'-tetrahydroxyaurone 4-O- $\beta$ -D-glucoside), (**2**) has also been found in this plant material.

Earlier work on the polyphenolic constituents of *Acacia dealbata* (Leguminosae) has led to the identification of 2 flavonol glycosides (myricetin 3-glucoside<sup>2</sup> and rutin<sup>3</sup>), a 5-deoxyflavonol (fisetin<sup>4</sup>), a leucoanthocyanidin (mollisacacidin<sup>5</sup>) and a flavanone glycoside (naringenin 5-O-diglucoside<sup>6</sup>). Moreover the presence of anthochlor pigments in the flowers of this plant has recently been shown by the isolation of 2 chalcone glycosides<sup>7</sup> (isosalipurposide and chalcononaringenin 2'-xyloside). The present study deals with the isolation of 2 further anthochlors (**1** and **2**) from the yellow flowers of *Acacia dealbata*.

**Material and methods.** For paper chromatography and TLC the solvent mixtures used, with their abbreviations, are as follows: A, 1-butanol-acetic acid-water (4:1:5, upper phase); B, acetic acid-water (5:95); C, 1-butanol-ethanol-water (4:1:2.2); D, ethyl acetate-butanone-formic acid-water (5:3:1:1); E, 1-butanol-pyridine-water (6:4:3); F, 1-butanol-acetic acid-ethyl ether-water (9:6:3:1); G, water; H, acetic acid-water (3:7); I, benzene-pyridine-formic acid (36:9:5); L, chloroform-methanol-butanone (70:10:6); M, benzene-methanol-butanone (3:1:1); N, chloroform-ethyl acetate (1:1); O, benzene-acetone (9:1.5); P, phenol saturated with water.

Fresh flowers (50 g) of *Acacia dealbata* Link (collected in Catania) were extracted 3 times with boiling 95% ethanol; the combined extracts were filtered, concentrated to a small volume in vacuo, re-filtered and evaporated to dryness in vacuo giving an orange-coloured residue (ca. 12 g). Anthochlor pigments **1** (circa 10 mg) and **2** (circa 25 mg) were isolated by preparative chromatography on Whatman 3MM paper in solvent A. Bands were cut off, eluted with 70% ethanol, concentrated in vacuo and rechromatographed in solvents B and C; further purification was obtained by preparative SiO<sub>2</sub> TLC in solvent D.

The UV-spectrum of pigment **1** ( $R_f$  values on Whatman No.1 paper: 0.69 in solvent A; 0.77 in solvent C; 0.30 in solvent G; 0.70 in solvent P) showed  $\lambda_{\text{max}}^{\text{MeOH}}$  (nm) 310 (sh), 368; +NaOMe 245 (sh), 340 (sh), 433 (increase in intensity); +AlCl<sub>3</sub> 260, 327, 420; +AlCl<sub>3</sub>/HCl 260, 320, 418; +NaOAc 260 (sh), 340 (sh), 395. These spectral properties are consistent<sup>8</sup> with pigment **1** (color reactions: brown to orange in UV+NH<sub>3</sub>) being a polyhydroxychalcone with free hydroxyl groups at positions 4 and 2'. Partial acid hydrolysis with 10% acetic acid (3.5 h under reflux), total acid hydrolysis with 2N HCl (2 h at 100 °C) and treatment with  $\beta$ -glucosidase gave D-glucose and homoeriodictyol (5,7,4'-trihydroxy-3'-methoxyflavanone). The sugar was identified by paper chromatography (solvents A and E), SiO<sub>2</sub> TLC (solvent F) and by the use of glucose oxidase; homoeriodictyol was identified by paper co-chromatography with authentic sample (solvents A, G and H), SiO<sub>2</sub> TLC (solvent I), polyamide TLC (solvents L and M) and UV-spectral analysis with shift reagents<sup>8</sup>. On methylation (methyl iodide in dimethylformamide in the presence of silver oxide), this chalcone glucoside gave a methyl ether,  $\lambda_{\text{max}}^{\text{MeOH}}$  350 nm, which on acid hydrolysis with 0.3 N HCl (4 h under reflux) isomerized to a flavanone since it gave eriodictyol tetramethyl ether (5,7,3',4'-tetramethoxyflavanone) and 2,3,4,6-tetra-O-methyl-D-glucose. The methylated sugar was identified by SiO<sub>2</sub> TLC (solvent N) and paper co-chromatography according to Petek<sup>9</sup>; eriodictyol tetramethyl ether was identified by UV-spectroscopy and alkaline degradation<sup>10</sup> to give di-O-methylphloroglucinol and 3,4-dimethoxycinnamic acid which were identified by SiO<sub>2</sub> TLC (solvent O). Thus pigment **1** must be 4,2',4',6'-tetrahydroxy-3-methoxychalcone 2'-O- $\beta$ -D-glucoside, a name which derives from its isomeric flavanone. Pigment **2** was identified as cernuoside (4,6,3',4'-tetrahydroxyaurone 4-O- $\beta$ -D-glucoside) by UV-spectral analysis with usual shift reagents<sup>8</sup>, paper co-chromatography with authentic material (solvents A, H and P) and SiO<sub>2</sub> TLC (solvent D).



This identification was confirmed by total acid hydrolysis, partial acid hydrolysis and treatment with  $\beta$ -glucosidase to give D-glucose and aureusidin (4,6,3',4'-tetrahydroxyaurone). D-glucose was identified as above; aureusidin was identified by paper co-chromatography with authentic sample (solvents A, H and P) and UV-spectral analysis with shift reagents<sup>8</sup>.

**Results and discussion.** Although several natural products based upon 3,4,2',4',6'-pentahydroxychalcone are known<sup>11</sup>, pigment **1** is the first chalcone whose substitution pattern is related to that of the flavanone homoeriodictyol. Cernuoside (**2**) is reported for the first time in the Leguminosae;

this anthochlor pigment was first isolated from *Oxalis cernua*<sup>12</sup> (Oxalidaceae) and later found in 4 species of Gesneriaceae (*Chirita micromusa*<sup>13</sup>, *Cyrtandra oblongifolia*<sup>14</sup>, *Didymocarpus malayanus*<sup>14</sup> and *Petrocosmea kerrii*<sup>15</sup>) and in the Plumbaginaceae (*Limonium bonduelli*<sup>13</sup> and *Limonium* cv. Gold Coast<sup>16</sup>). It is interesting to note that anthochlor pigments **1** and **2** as well as anthochlors (isosalipurposide and chalcononaringenin 2'-xyloside) previously isolated<sup>7</sup> from *Acacia dealbata* have phloroglucinol A-ring structures. The occurrence of 4 such pigments in this plant is quite exceptional since resorcinol based-A-ring anthochlors are considered a biochemical characteristic of the family Leguminosae<sup>11</sup>.

- 1 Acknowledgments. The author thanks Prof. H. Wagner, Institut für pharmazeutische Arzneimittellehre der Universität München, for a sample of homoeriodictyol, and Prof. J.B. Harborne, University of Reading, for a sample of aureusidin.
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## Starfish saponins VII. Structure of luzonicoside, a further steroidal cyclic glycoside from the pacific starfish *Echinaster luzonicus*<sup>1,2</sup>

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**Summary.** On the basis of comparative chemical and spectral data, the structure of the major saponin, luzonicoside, from the starfish *Echinaster luzonicus* has been elucidated as **2**. This is a further example of a novel class of steroidal cyclic glycoside from starfish of the genus *Echinaster*. Its structure includes a  $\Delta^7$ , 3 $\beta$ ,6 $\beta$ -dioxxygenated-23-oxosteroidal moiety, already found in the saponins of *Echinaster sepositus*, and a trisaccharide moiety,  $\beta$ -D-galactopyranosyl-(1  $\rightarrow$  2)  $\alpha$ -L-arabinopyranosyl-(1  $\rightarrow$  2)- $\beta$ -D-glucuronopyranosyl, bridging C-3 and C-6 of the steroid.

Recently we have elucidated<sup>6</sup> the structure of the major saponin, sepositoside A, from *Echinaster sepositus* as **1**. On very mild acid treatment (1N-HCl, r.t.) it gives the corresponding opened glycoside **3**, while it is hydrolyzed to its sugars and the 3 $\beta$ -hydroxy-5 $\alpha$ -cholesta-8,14-dien-23-one<sup>7</sup>, by prolonged acid treatment. The structures of the minor saponins from the same sources possess the same cyclic trisaccharide moiety bridging C-3 and C-6 of the steroid, and the differences reside in the steroidal side-chains, characterized by epoxide functionalities at C-22 and C-23 (part structures **5-7**)<sup>6</sup>.

In this paper we describe the discovery of a further example of this novel class of steroidal cyclic glycoside from a starfish of the same genus, *E. luzonicus*, collected near Nouméa, Nouvelle Calédonie. The extraction and isolation of the saponins has followed the same procedure used before<sup>6</sup>. Fresh animals (4 kg), collected in october 1979, were extracted with water and the extracts were lyophilized (290 g). After removal of fat materials by washing with CHCl<sub>3</sub>, the saponins were recovered from the aqueous solution by Amberlite XAD-2 and purified by silica gel

column chromatography followed by reverse phase HPLC<sup>9</sup> (C-18  $\mu$ -Bondapak; CH<sub>3</sub>OH:H<sub>2</sub>O, 55:45) to give 0.75 g of luzonicoside.

Luzonicoside (**2**) analyzed for C<sub>44</sub>H<sub>67</sub>O<sub>17</sub>Na, was levorotatory, [ $\alpha$ ]<sub>D</sub> -66° (H<sub>2</sub>O), and was indistinguishable from sepositoside A (**1**) in TLC and HPLC. On acid hydrolysis it yielded the 3 $\beta$ -hydroxy-5 $\alpha$ -cholesta-8,14-dien-23-one, but, unlike **1**, D-galactose, L-arabinose and D-glucuronic acid. In the 270-MHz NMR-spectrum (DMSO) it showed a broad signal at  $\delta$ 5.47 ( $W_{1/2}$  = 12 Hz) corresponding to the olefinic proton at C-7, 2 methyl singlets at  $\delta$ 0.556 and 0.825 corresponding to CH<sub>3</sub>-18 and CH<sub>3</sub>-19, respectively and methyl doublets at  $\delta$ 0.85 and 0.83 for CH<sub>3</sub>-21, -26 and -27. The same signals were also observed in the spectrum of **1**; the spectrum of **2** also contained 3 doublets assigned to the anomeric protons at  $\delta$ 4.32 ( $J$  = 7.5 Hz), 4.65 ( $J$  = 7.5 Hz) and 5.05 ( $J$  = 7.0 Hz). Similarly the comparison of the <sup>13</sup>C-NMR spectrum of **2** with that of the known **1** indicated that the steroidal parts of the molecules were identical (all resonances associated with the steroidal carbons had identical